

Electrosynthesis of 4,4-Dimethyl-2-ethoxycarbonyl-5-fluoro-3-thiolanones: Highly Potent Human Type II PLA₂ Inhibitors¹

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Abstract: Anodic monofluorination of 2-methyl- and 2-benzyl-4,4-dimethyl-2-ethoxy-carbonyl-3-thiolanones **1** and **2** was successfully performed to provide the corresponding 5-fluorinated products **3** and **4**, respectively in good yields. The stereoisomeric mixture of **4** was found to possess comparable or even stronger *in vitro* human type II phospholipase A₂ inhibitory activity compared with the known inhibitor, manoalide, and the cis isomer of **4** exhibited higher activity than the trans isomer.

Incorporation of fluorine(s) into organic molecules sometimes enhances their biological activities.² In fact, fluoroketones, fluorobenzylamines, and fluorinated 4-aminobutyrophenones have been reported to be potent inhibitors of phospholipase A₂.³ In order to develop such pharmaceutically active compounds, selective direct fluorination of organic molecules has been increasingly important. However, the fluorination is not always straightforward because conventional direct fluorination usually requires dangerous, troublesome, or costly fluorinating reagents.⁴ Recently, we have demonstrated that anodic fluorination was much superior to conventional chemical methods.⁵

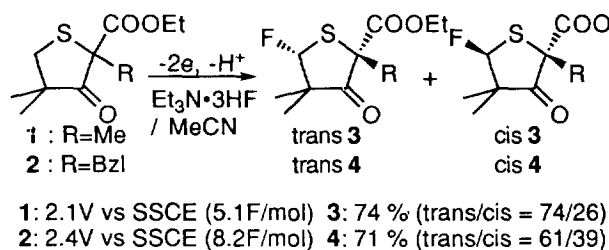
In this paper, we report anodic monofluorination of 2-methyl- and 2-benzyl-4,4-dimethyl-2-ethoxycarbonyl-3-thiolanones **1** and **2** together with human type II phospholipase A₂ (PLA₂) inhibitory activities of the monofluorinated products. Anodic fluorination of 3-thiolanone derivatives **1** and **2** were performed as follows. Constant potential electrolysis was carried out at a platinum cathode and anode (2x2cm) in 0.37M Et₃N•3HF/MeCN (15mL) containing 1.5mmol of **1** or **2** in a cylindrical one-compartment glass cell. After the starting material was completely consumed, the electrolysis solution was passed through a short column of silica gel (CH₂Cl₂). The products **3** and **4** were isolated by silica gel column chromatography (hexane, ether=2:1~4:1). The cis and trans stereoisomers of **4**⁶ could be separated by TLC (hexane, ether=3:1) and their structures were determined by X-ray crystallographic analysis. On the other hand, attempts to separate the isomers of **3**⁷ were unsuccessful until now although assignment of ¹⁹F NMR spectrum of each isomer could be done.

As shown in Scheme 1, desired monofluorinated products **3** and **4** were obtained in rather good yields. In both cases, trans isomers were major products. For each of these monofluorinated products and starting materials, the *in vitro* human type II phospholipase A₂ inhibitory activity was assayed.⁸

Table 1 shows the results for **3** and cis and trans isomers of **4** together with the known PLA₂

inhibitor, manoalide⁹, as the reference compound. A comparison of IC₅₀ values clearly indicates the potential of the monofluorinated heterocycles **4** as an anti-inflammatory substance, which showed more effective nature than manoalide in inhibition of human type II phospholipase A₂. In sharp contrast to **4**, starting nonfluorinated **2** and fluorinated **3** did not possess such activity. Furthermore, it was also found that the *cis* isomer of **4** showed higher activity than the *trans* isomer.

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(Scheme 1)

Table 1. Human Type II Phospholipase A₂ Inhibitory Activity *in vitro*⁸

Compound	IC ₅₀ (μg/mL)
4 (<i>cis/trans</i> mixture)	0.20
<i>cis</i> - 4	0.21
<i>trans</i> - 4	1.27
manoalide	0.34

References and Notes

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- cis*-**4**: colorless oil. ¹⁹F NMR (CDCl₃) δ -69.24 (dd, J_{H,F}=56.44, 2.48 Hz). ¹H NMR (CDCl₃) δ 1.12 (d, 3H, C(CH₃)₂, J=2.97 Hz), 1.24 (t, 3H, CH₂CH₃, J=6.93 Hz), 1.27 (s, 3H, C(CH₃)₂), 3.16 (d, 1H, CH₂Ph, J=13.53 Hz), 3.52 (d, 1H, CH₂Ph, J=13.86 Hz), 4.19 (q, 2H, CH₂CH₃, J=6.93 Hz), 5.62 (d, 1H, CHF, J=57.07 Hz), 7.22-7.26 (m, 5H, aromatic H).
trans-**4**: colorless solid, mp 83.5-84.0 °C. ¹⁹F NMR (CDCl₃) δ -69.88 (d, J_{H,F}=56.41 Hz). ¹H NMR (CDCl₃) δ 0.41 (d, 3H, C(CH₃)₂, J=1.32 Hz), 1.24-1.26 (m, 6H, CH₂CH₃, & C(CH₃)₂), 3.27 (d, 1H, CH₂Ph, J=14.18 Hz), 3.51 (d, 1H, CH₂Ph, J=14.52 Hz), 4.12-4.33 (q, 2H, CH₂CH₃, J=6.93 Hz), 5.49 (d, 1H, CHF, J=56.41 Hz), 7.12-7.28 (m, 5H, aromatic H). IR (KBr, neat) 3036, 2982, 2934, 2903, 1743, 1728, 1605, 1493, 1472, 1456, 1387, 1366, 1333, 1275, 1232, 1205, 1145, 1119, 1080, 1046, 1026, 999, 966, 924, 860, 808, 758, 704 cm⁻¹. MS: m/z 310 (M⁺), 262, 203, 135, 91. HRMS: calcd for C₁₀H₁₉FO₃S 310.1037, found 310.1029.
- cis/trans* mixture: colorless oil. ¹⁹F NMR (CDCl₃) *trans* isomer δ -67.2 (d, J_{H,F}=58.0 Hz); *cis* isomer δ -71.2 (d, J_{H,F}=56.0 Hz). ¹H NMR (CDCl₃) δ 1.23-1.37 (m, 9H, CH₂CH₃, & C(CH₃)₂), 1.67 (s, 3H, CH₃; *cis* isomer), 1.76 (s, 3H, CH₃; *trans* isomer), 4.16-4.24 (m, 2H, CH₂CH₃), 5.27 (d, 1H, CHF, J=56.08 Hz), 5.29 (d, 1H, CHF, J=55.75 Hz). MS m/s: *trans* isomer 234 (M⁺), 161, 59; *cis* isomer 234 (M⁺), 161, 59.
- The inhibitory activity was assayed with 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphoethanolamine as a substrate, according to the method described by Tojo et al. with a slight modification: Tojo, H.; Ono, T.; Okamoto, M. *Methods in Enzymology*, **1991**, *197*, 390.
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